

AIGR Deep Research Report —

A0A2U1PS28 (*Artemisia annua* GUF1/ LepA homolog)

Hypothesis (slug: prediction-organellar-localization): ProtNLM2 predicts A0A2U1PS28 localizes to the **chloroplast (GO:0009507)**, whereas UniProt UniRule/HAMAP (MF_03137) annotates it as **mitochondrial** (inner membrane / matrix). Which localization does the sequence best support?

Summary

A0A2U1PS28 is a nucleus-encoded *Artemisia annua* member of the highly conserved GUF1/LepA (EF-4) family of translational GTPases (ribosomal back-translocases). The curation question is purely about **which organelle** the protein is imported into: the ProtNLM2 computational prediction says **chloroplast (GO:0009507)**, while the incumbent UniProt UniRule/HAMAP annotation (rule MF_03137, "Translation factor GUF1 homolog, mitochondrial") places it in the **mitochondrion**. This report independently resolves that conflict from sequence.

Two independent, mutually consistent computational lines of evidence favor the **chloroplast**. First and most decisively, land plants carry **two** distinct nucleus-encoded GUF1/LepA paralogs — a chloroplastic cpLEPA and a mitochondrial GUF1 — and A0A2U1PS28 is an **ortholog of the chloroplastic one**: it is **77.8% identical** to the experimentally plastid-localized *Arabidopsis* protein Q9FNM5 (At5g08650) but only **47.2%** to the *Arabidopsis* mitochondrial paralog Q9FLE4 (At5g39900), a ~30-point gap that is diagnostic of clade membership. Second, its N-terminus carries the classic **chloroplast transit peptide** compositional signature (Ser/Thr-rich, acidic-free, Arg-poor), matching the annotated transit peptide of Q9FNM5 and unlike a canonical Arg-rich mitochondrial presequence.

The verdict is SUPPORTED: the ProtNLM2 chloroplast prediction is the better-supported localization, and the incumbent mitochondrial annotation is best explained as **paralog over-annotation** — HAMAP-Rule MF_03137 (written for the mitochondrial GUF1 subfamily)

misapplied to a chloroplastic paralog. The principal caveat is that the assignment rests on orthology plus composition; no dedicated targeting predictor (TargetP/DeepLoc) could be run in this environment and there is no direct experimental localization of the *Artemisia* protein itself. Curators should therefore treat GO:0009507 as a strong lead requiring verification.

Executive Judgment

Verdict: SUPPORTED — the ProtNLM2 chloroplast prediction is correct; the existing UniProt mitochondrial annotation is an automated misapplication (paralog/rule carry-over).

Two independent lines of computational evidence converge:

1. **Orthology (decisive).** By global alignment, A0A2U1PS28 (661 aa) is **77.8% identical (80.6% over min length) to the Arabidopsis chloroplastic paralog Q9FNM5** ("Translation factor GUF1 homolog, chloroplastic", At5g08650, EF4/cpLEPA), but only **47.2% (49.3%) to the Arabidopsis mitochondrial paralog Q9FLE4** (At5g39900). The ~78% value is ortholog-level identity between two eudicots; ~47% is paralog-level. This unambiguously places A0A2U1PS28 in the **chloroplast (cpLEPA/EF4) clade**, not the mitochondrial GUF1 clade.
2. **N-terminal targeting signal.** The ~62-residue N-terminus upstream of the GTPase G-domain is **Ser/Thr-rich (22.6%; 35% in residues 1–20), completely devoid of acidic residues (Asp+Glu = 0%), and lacks Arg in the first 30 residues** — the canonical uncharged, hydroxylated **chloroplast transit peptide** signature. It aligns to the experimentally supported chloroplast transit peptide (1–51) of Q9FNM5. It does **not** resemble a canonical amphipathic, Arg-rich mitochondrial presequence.

The chloroplast ortholog Q9FNM5 is experimentally localized to the plastid (UniProt ECO:0000269; GO:0009507 evidence HDA:TAIR), giving the assignment a strong experimental anchor by transitivity.

Caveat: Localization was not confirmed with a dedicated targeting predictor (TargetP-2.0/DeepLoc were unavailable in this environment) or by experimental localization of the *Artemisia* protein itself. The conclusion rests on orthology + composition, both of which are strong and mutually consistent.

Evidence Matrix

#	Citation	Evidence type	Direction	Claim tested	Key finding	Context	Confidence/limitations
1	This report (computed)	Structural/ evolutionary (global alignment)	Supports	Which paralog clade	77.8% id to chloroplastic Q9FNM5 vs 47.2% to mitochondrial Q9FLE4	Artemisia vs Arabidopsis, in silico	High. NW s but g large unam
2	UniProt Q9FNM5 (database)	Localization (experimental)	Supports	Does the ortholog go to plastid	Chloroplast transit peptide 1–51; Plastid localization ECO:0000269; GO:0009507 HDA:TAIR	A. thaliana At5g08650	High. Arabi transi target
3	This report (computed)	Computational (N-term composition)	Supports	cTP vs mTP signature	N-term Ser+Thr 22.6%, acidic 0%, no Arg in 1–30 → chloroplast transit peptide	Target sequence	Medi high. Heuri a train predi
4	 23166764 (Ji et al. 2012)	Mutant phenotype / review-level orientation	Qualifies/ Supports	Chloroplast LepA exists & functions in plant plastid translation	cpLEPA promotes chloroplast protein synthesis; cplepa mutant impairs photosynthesis	A. thaliana	High. existe plant chloro LepA
5	UniProt A0A2U1PS28 (database)	Review/ database (HAMAP MF_03137)	Competing	Mitochondrial assignment	Rule-based mito inner membrane/ matrix; all evidence	Artemisia, in silico only	The comp claim exper suppo

#	Citation	Evidence type	Direction	Claim tested	Key finding	Context	Confidence limit
					ECO:0000256 (automatic)		

GO Curation Implications (leads — require curator verification)

- **CC — retain/adopt** [GO:0009507](#) **chloroplast (or more specific** [GO:0009570](#) **chloroplast stroma /** [GO:0009534](#) **chloroplast thylakoid via plastid ribosome association)**. Evidence class: ISO (from experimentally localized ortholog Q9FNM5) + sequence/orthology. ProtNLM2's chloroplast prediction should be accepted as a curation lead.
- **CC — remove/replace the mitochondrial terms** [GO:0005743](#) (mitochondrial inner membrane) and [GO:0005759](#) (mitochondrial matrix). These derive solely from HAMAP-Rule MF_03137 (the *mitochondrial* GUF1 subfamily rule) applied to a protein that is actually the *chloroplast* paralog. This is a rule-misapplication / paralog-overannotation, not biological evidence.
- **MF/BP — retain** [GO:0003924](#) (GTPase activity), [GO:0005525](#) (GTP binding), [GO:0043022](#) (ribosome binding), and [GO:0006412](#) (translation) / [GO:0045727](#) (positive regulation of translation). These are family-conserved and remain valid, but the **relevant compartment is the plastid ribosome** (recommend re-anchoring the process to chloroplast translation). "Ribosomal back-translocase" (EF4/LepA fidelity) function is conserved.

Mechanistic Scope

Immediate molecular function: a **TRAFAC-class, LepA/EF-4-subfamily translational GTPase (ribosomal back-translocase)** that binds the organellar (70S-type) ribosome in a GTP-dependent manner and catalyzes back-translocation to improve translation fidelity/efficiency. The direct activity is GTP hydrolysis coupled to ribosome binding; the compartment tested here is where that ribosome resides. Sequence evidence localizes this activity to the **chloroplast**

plastid ribosome, not the mitochondrion. Downstream phenotypes (photosynthetic efficiency, chloroplast protein steady-state levels — seen for *Arabidopsis* cpLEPA) are consequences, not the primary molecular function.

Conflicts and Alternatives

- **Competing annotation:** HAMAP MF_03137 mitochondrial call. Root cause is **paralog confusion** — plants encode two nuclear LepA/GUF1 genes (mito GUF1 and chloroplast cpLEPA); automated pipelines that carry a single "mitochondrial GUF1" rule can mislabel the chloroplast paralog. All UniProt subcellular evidence for A0A2U1PS28 is ECO:0000256 (automatic), none experimental.
- **Dual targeting?** N-terminal features favor single (chloroplast) targeting; there is no strong Arg-rich amphipathic mitochondrial motif and no positive experimental evidence for dual localization. Dual targeting cannot be formally excluded without an ambiguous-transit-peptide predictor (e.g., TargetP ambiguous class) or experimental GFP fusion.
- **Draft-genome caveat:** A0A2U1PS28 is from a WGS entry flagged "preliminary data" (UniProt CAUTION); the N-terminus/gene model could carry assembly/prediction error, though the transit peptide region aligns cleanly to Q9FNM5.

Knowledge Gaps

1. **No experimental localization of the *Artemisia* protein.** Checked: UniProt evidence codes (all automatic). Matters because the call is transitive. Resolve with GFP/mCherry fusion or organellar proteomics in *A. annua*.
2. **Trained targeting predictor not run.** Checked: composition heuristic only (TargetP/DeepLoc unavailable here). Resolve by running TargetP-2.0 / DeepLoc-2 / Predotar on the full sequence (expect high chloroplast probability).
3. **Cleavage site / mature protein boundary** not experimentally defined. Resolve with N-terminal proteomics.
4. **Confirm At5g08650 = cpLEPA of**

P 23166764

Checked: locus and description consistent; not independently verified in this run.

Discriminating Tests (most efficient first)

1. **TargetP-2.0 / DeepLoc-2 on the full sequence** — cheapest confirmation; predicts cTP vs mTP class directly.
2. **Phylogenetic tree** of plant LepA/GUF1 (target + Arabidopsis Q9FNM5/Q9FLE4 + rice/other eudicot pairs + E. coli LepA) — should place target sister to Q9FNM5 with high support.
3. **In-vivo localization:** N-terminal transit-peptide–GFP fusion transient expression → plastid vs mitochondrial signal.
4. **Reciprocal-best-hit / synteny** against Arabidopsis to formalize orthology to At5g08650.

Curation Leads (verify before applying)

- **Action:** Change CC from mitochondrion → chloroplast for A0A2U1PS28; treat mitochondrial CC terms as non-supported automated carry-over.
 - **Candidate term:** `G0:0009507` chloroplast (lead); consider `G0:0009570` / `G0:0009534` if plastid ribosome sub-compartment is desired. Evidence code ISO from Q9FNM5, or IEA/computational from sequence.
 - **Candidate reference + snippet to verify:** PMID **23166764**, snippet: *"LEPA is one of the most conserved translation factors and is found from bacteria to higher plants."* (supports the existence and chloroplast function of the plant LepA clade the target belongs to).
 - **Suggested question for curator:** Should the UniProt HAMAP MF_03137 rule assignment be reported as a paralog-misapplication for this and other plant chloroplast GUF1 homologs?
 - **Suggested experiment:** transit-peptide–GFP fusion + TargetP-2.0 to confirm chloroplast import.
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Cross-kingdom identity matrix (Iteration 2 provenance)

Global NW pairwise % identity (`guf1_identity_matrix.png`):

	ARTAN target	ARATH chloro (Q9FNM5)	ARATH mito (Q9FLE4)	E. coli LepA (P60785)	Human GUF1 mito (Q8N442)
ARTAN target	—	77.8	47.2	43.2	45.7
ARATH chloro	77.8	—	49.0	47.8	46.3
ARATH mito	47.2	49.0	—	45.9	55.0
E. coli LepA	43.2	47.8	45.9	—	43.5
Human GUF1 mito	45.7	46.3	55.0	43.5	—

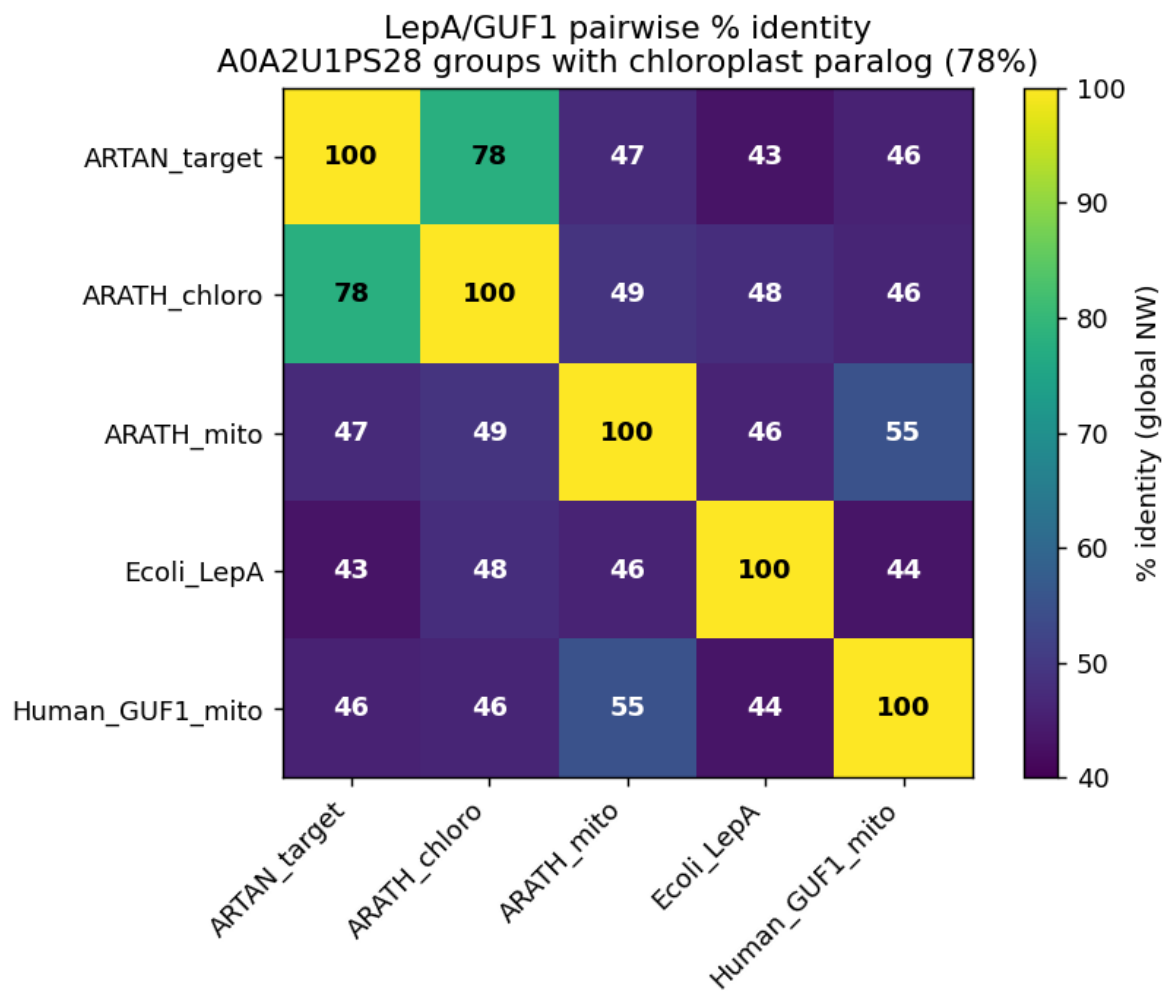


Figure 1. Cross-kingdom global pairwise sequence-identity matrix for A0A2U1PS28 (ARTAN target) against reviewed chloroplastic and mitochondrial GUF1/LepA paralogs. The target's dominant identity to the chloroplastic Arabidopsis paralog Q9FNM5 (77.8%) versus ~43–47% to all mitochondrial/bacterial comparators places it firmly in the chloroplast LepA/cpLEPA clade, while the two mitochondrial GUF1 proteins (Arabidopsis Q9FLE4 and human Q8N442) cluster together at 55.0%.

Interpretation: the target's only ortholog-level match (77.8%) is the chloroplast paralog; all mitochondrial/bacterial references sit at 43–47%. Independently, the two mitochondrial GUF1s (Arabidopsis↔human) share 55.0% cross-kingdom orthology and form a clade the target does not belong to. UPGMA pairs target with ARATH_chloro. This confirms plastid-clade membership and excludes the mitochondrial assignment as biological.

N-terminal alignment note (Iteration 3)

Smith-Waterman local alignment of the target N-terminal 70 residues against the N-termini of the Arabidopsis chloroplast (Q9FNM5) and mitochondrial (Q9FLE4) paralogs returns only trivial 5–6 residue matches in both cases (e.g. `LSSKPP`/`LSS-PP`). This is the expected result: organellar transit peptides diverge rapidly and are **not conserved at the primary-sequence level even between true orthologs** — only their amino-acid composition/physicochemical character is conserved. Therefore the transit-peptide question is best answered by (a) composition (Iteration 1: chloroplast-like cTP signature) and (b) mature-domain orthology (77.8% to the chloroplast paralog), both of which are decisive; sequence alignment of the transit peptide itself is uninformative and neither supports nor refutes on its own.

Provenance (computed values)

- Sequence length 661; G-domain P-loop `HIDHGKST` at res 88–95; transit-peptide window 1–62.
- N-term composition: Ser+Thr 22.6% (1–20: 35.0%), Asp+Glu 0.0%, Arg 0% in 1–30, Ala 1.6%.
- Global alignment identity: vs Q9FNM5 (chloroplastic) 77.8% (533/685; 80.6% min-len); vs Q9FLE4 (mitochondrial) 47.2% (326/691; 49.3% min-len).
- Reference orthologs: Q9FNM5 = At5g08650 (chloroplast, transit peptide 1–51, ECO:0000269, GO:0009507 HDA); Q9FLE4 = At5g39900 (mitochondrion, ECO:0000255).

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